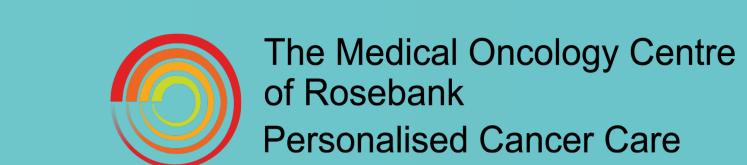




1150P - Transforming growth factor-beta-1 and soluble co-inhibitory immune checkpoints as putative drivers of immune suppression in advanced basal cell carcinoma



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Background

- ▶ Basal cell carcinoma (BCC) is the most common malignancy, comprising about 75 % of all cases of skin cancer, and the incidence is rising^{1,2}.
- ▶ BCC rarely metastasizes and the mortality rate is low; however, the disease is associated with substantial morbidity³. ▶ The hedgehog intracellular signalling pathway regulates cell growth, and aberrant activation of
- this pathway leads to BCC development³. The hedgehog inhibitors vismodegib and sonidegib are currently approved for systemic therapy of BCC in Europe^{3,4,5}. ▶ Hedgehog-dependent tumors are characterized by increased infiltration or the presence of

suppressive immune cells, such as M2-like tumor-associated macrophages (M2-TAMs), myeloid-

- derived suppressor cells (MDSCs), regulatory T (Treg) cells, and cancer-associated fibroblasts ▶ BCC is associated with increased numbers of regulatory cells (Tregs) and a CAF-induced
- immunosuppressive microenvironment¹¹⁻¹⁴. ▶ Checkpoint proteins are critical for maintaining self-tolerance and modulating the immune
- responses of effector cells in normal tissues to minimize tissue damage. These proteins also modulate the immune infiltrates in the tumor microenvironment (TME). Cancer cells exploit the up-regulation or down-regulation of these proteins to evade the anti-tumor immune response^{15,16}. > Soluble forms of immune checkpoint molecules (ICMs) have recently been identified and can be
- measured in human plasma; however, their biological and clinical significance remains essentially unknown^{17,18}. Co-inhibitory immune checkpoint proteins are primarily involved in promoting inhibitory cell-cell interactions in adaptive immunity, especially tumor immunity.
- ▶ The soluble cell-free variants of these molecules are detectable in the circulation of cancer patients where they retain immunosuppressive activity.
- Little is known about the systemic levels of these soluble co-inhibitory and co-stimulatory immune checkpoints in patients with various subtypes of basal cell carcinoma (BCC), which is the most invasive and treatment-resistant type of this most commonly occurring malignancy.
- ▶ Our previous research found significantly elevated levels of PD-1, PDL-1, CTLA-4, TIM-3, and LAG-3 in BCC patients and the current study was undertaken to investigate 16 ICM proteins as well as RANTES, FAP, TGF-β1 and arginase.

Methods

- ▶ The study population consisted of a total of 40 South African patients (12F:28M; mean age ±SD: 69.1 ± 11.1 years) with advanced BCC attending the Dermatology Screening Clinic at Steve Biko Academic Hospital, Pretoria, South Africa.
- ▶ The cohort was compared with a group of control patients (n=20).
- ▶ The circulating levels of 17 immune checkpoint-related proteins panel (B- and T-lymphocyte attenuator (BTLA), Glucocorticoid-Induced TNFR-Related protein (GITR), GITR-ligand (GITRL), Herpes Virus Entry Mediator (HVEM), Lymphocyte activation gene-3 (LAG-3), PD-1, PD-L1, PD-L2, T cell immunoglobulin-3 (TIM-3), CD27, CD28, CD80, CD86, CD40, ICOS, TLR-2, and CTLA-4) were profiled in BCC patients (patient characteristics are summarized in table 1) and compared to those of 20 healthy controls.
- ▶ Additionally, we measured plasma levels of arginase, CCL5 (RANTES), TGF-β1 and fibroblast associated protein (FAP).
- ▶ A combination of multiplex bead array, laser nephelometry and ELISA technologies were used. ▶ Ethics approval was granted by the Research Ethics Committee of the Faculty of Health Sciences,

Statistical Analysis

- ▶ The primary hypothesis was that that there was a significant difference in the plasma levels of soluble co-inhibitory immune checkpoints between BCC patients and healthy controls.
- Descriptive statistics were used to tabulate patient characteristics.
- The Mann Whitney U-test was used to compare levels of the various test biomarkers between BCC patients and healthy controls. The area under the ROC curve (AUC) was used as a measure of discriminatory ability for the
- biomarkers. The Youden index, a summary measure of the ROC curve, was used as an agnostic method for choosing an optimal cut-off value on the biomarker value to illustrate potential clinical usefulness.
- A correlation matrix report was used to identify correlations between variables (or subsets of variables) within the subset, using Spearman *p*-values to define significance.
- A p-value of less than .05 was considered statistically significant.
- ▶ NCSS 2021 software for Windows (USA) was used for statistical analyses.

Results

Table 1. Numbers of patients with distinct Table 2. Numbers of patients with basal clinical types of basal cell carcinoma (BCC). cell carcinomas at distinct anatomical

Clinical subtype of BCC	;	Anatomical site		
Adenoid	(n=1)*	Cheek	(n=3)*,+	
Basosquamous	(n=3)	Chest	(n=2)	
	(n=22)	Ear	(n=4)	
Infiltrating		Forearm	(n=4)	
Infiltrating with squamous differentiation	(n=4)	Forehead	(n=2)	
Keratotic	(n=1)	Lower limb	(n=5)	
Micronodular	(n=2)	Neck	(n=2)	
Nodular	(n=5)	Nose	(n=13)°	
Noutiai	(11–3)	Shoulder	(n=1)	
Pigmented	(n=1)+	Temple	(n=2)	
Superficial	(n=1)°	Upper anterior chest	(n=2)	

*Numbers of patients are shown in parenthesis; *African patient; °Asian patient

Table 3. Comparison of the systemic concentrations of co-inhibitory, and costimulatory soluble immune checkpoint proteins in patients with advanced basal cell carcinoma and control participants.

			BCC (n=40)	Controls (n=20)		
	ICM		Median pg/ml (95%Cl)	Median pg/ml (95%Cl)	p value	
Co-stimulatory	CD27	UP	3360,665 (2363,64 - 4970,73)	1410,54 (1259,16 - 2172,74)	0,0002	
	CD28	UP	17047,05 (8487,16 - 30677,1)	11314,17 (7236,45 - 14883,36)	0,2523	
	CD40	UP	1308,5 (968,17 - 1779,77)	1222,255 (769,43 - 1349,26)	0,4148	
	ICOS	UP	15359,79 (7591,11 - 20308,75)	12902,86 (7980,59 - 15316,53)	0,3428	
o-stim	GITR	UP	1217,4 (664,31 - 1795,54)	698,205 (228,01 - 1222,24)	0,0538	
ŏ	GITRL	UP	2527,32 (1470,48 - 3599,4)	2107,325 (1784,1 - 2724,34)	0,3799	
	CD86	UP	2215,865 (793,93 - 3292,67)	1636,65 (781,54 - 2144,3)	0,2427	
	CD80	UP	1450,26 (863,6 - 2161,26)	1212,29 (781,71 - 1590,1)	0,3428	
CO-inhibitory CTLA TIM-	PD-1	UP	10978,21 (5714,49 - 14351,17)	2524,69 (1832,95 - 3038,34)	0,0000	
	PD-L1	UP	1740,25 (773,982 - 1980,649)	228,67 (139,61 - 274,66)	0,0000	
	PD-L2	UP	14705,27 (13102,68 - 16375,87)	12008,07 (10670,4 - 14023,9)	0,0011	
	CTLA-4	UP	744,92 (422,08 - 1129,16)	126,49 (56,24 - 241,25)	0,0000	
	TIM-3	UP	7519,74 (6619,886 - 8157,926)	12008,07 (10670,4 - 14023,9)	0,0000	
	LAG-3	UP	388288,90 (243248,3 - 540480,6)	11106,96 (6595,67 - 15093,31)	0,0000	
	BTLA	DOWN	12284,97 (8754,07 - 19151,59)	25439,74 (17274,69 - 32427,56)	0,0061	
TLF	TLR-2	UP	17696,28 (10473,49 - 24211,18)	15731,88 (12262,72 - 19913,19)	0,6437	
Dual	HVEM	UP	2052,45 (1894,5 - 2317,55)	1299,11 (1263,46 - 1458,94)	0,0000	
Other	Arginase		25,52 (25,52 - 29,8505)	25,52 (25,52 - 72,15)	0,2897	
	RANTES	UP	131,46 (97,25 - 174,9144)	90,83 (70,78 - 148,71)	0,2097	
	TGF-β1	UP	7,54 (4,549417 - 10,79543)	5,83 (4,18 - 6,83)	0,1469	
	FAP	UP	115,67 (94,02 - 130,19)	109,04 (70,83 - 127,33)	0,2425	

Table 4. ROC curve cut-off values (using Youden Index) and AUC (95% CI) for immune

Soluble Immune Checkpoint Molecule (pg/mL) Cut-off point (pg/mL) Sensitivity (TNR) % P ≤							
CD28 0,591 ≥ 20005,17 48 75 0,1066 CD40 0,565 ≥ 1701,52 40 90 0,1971 ICOS 0,576 ≥ 17225,11 48 85 0,1505 GITR 0,654 ≥ 2001,53 33 100 0,0158 GITR 0,570 ≥ 4107,31 33 100 0,1692 CD86 0,593 ≥ 2609,66 45 90 0,0998 CD80 0,576 ≥ 2200,90 33 100 0,1491 PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542	Check	point Molecule	AUC (CI 95%)				p≤
CD40 0,565 ≥ 1701,52 40 90 0,1971 ICOS 0,576 ≥ 17225,11 48 85 0,1505 GITR 0,654 ≥ 2001,53 33 100 0,0158 GITRL 0,570 ≥ 4107,31 33 100 0,1692 CD86 0,593 ≥ 2609,66 45 90 0,0998 CD80 0,576 ≥ 2200,90 33 100 0,1491 PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 LAG-3 1,000 ≥ 3774,69 98 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542 TGF-β1 0,616 2 8,21 43 100 0,0542 TGF-β1	ılatory	CD27	0,204	≤ 2989,38	40	100	1,0000
ICOS 0,576 ≥ 17225,11 48 85 0,1505 GITR 0,654 ≥ 2001,53 33 100 0,0158 GITR 0,570 ≥ 4107,31 33 100 0,1692 CD86 0,593 ≥ 2609,66 45 90 0,0998 CD80 0,576 ≥ 2200,90 33 100 0,1491 PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542 TGF-β1 0,616 2 8,21 43 100 0,0542 TGF-β1		CD28	0,591	≥ 20005,17	48	75	0,1066
CD86 0,593 ≥ 2609,66 45 90 0,0998 CD80 0,576 ≥ 2200,90 33 100 0,1491 PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,00542		CD40	0,565	≥ 1701,52	40	90	0,1971
CD86 0,593 ≥ 2609,66 45 90 0,0998 CD80 0,576 ≥ 2200,90 33 100 0,1491 PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,00542		ICOS	0,576	≥ 17225,11	48	85	0,1505
CD86 0,593 ≥ 2609,66 45 90 0,0998 CD80 0,576 ≥ 2200,90 33 100 0,1491 PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,00542	-stim	GITR	0,654	≥ 2001,53	33	100	0,0158
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	ပိ	GITRL	0,570	≥ 4107,31	33	100	0,1692
PD-1 0,874 ≥ 4849,52 73 95 0,0000 PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,00542		CD86	0,593	≥ 2609,66	45	90	0,0998
PD-L1 0,926 ≥ 404,54 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542		CD80	0,576	≥ 2200,90	33	100	0,1491
PD-L1 ≥ 404,34 89 95 0,0000 PD-L2 0,761 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542	, and	PD-1	0,874	≥ 4849,52	73	95	0,0000
PD-L2 ≥ 11788,96 90 50 0,0000 CTLA-4 0,889 ≥ 292,80 73 95 0,0000 TIM-3 0,996 ≥ 3774,69 98 100 0,0000 LAG-3 1,000 ≥ 33502,53 100 100 0,0000 BTLA 0,281 ≥ 15585,46 43 20 0,9991 TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542		PD-L1	0,926	≥ 404,54	89	95	0,0000
LAG-3		PD-L2	0,761	≥ 11788,96	90	50	0,0000
LAG-3	hibit	CTLA-4	0,889	≥ 292,80	73	95	0,0000
LAG-3	Co-in	TIM-3	0,996	≥ 3774,69	98	100	0,0000
TLR-2 0,537 ≥ 22486,86 40 90 0,3081 HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542		LAG-3	1,000	≥ 33502,53	100	100	0,0000
Position HVEM 0,916 ≥ 1524,59 90 90 0,0000 Arginase 0,420 ≥ 36,62 23 55 0,8537 RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542		BTLA	0,281	≥ 15585,46	43	20	0,9991
Arginase 0,420 \geq 36,62 23 55 0,8537 RANTES 0,600 \geq 198,72 33 95 0,0861 TGF-β1 0,616 \geq 8,21 43 100 0,0542	Dual	TLR-2	0,537	≥ 22486,86	40	90	0,3081
RANTES 0,600 ≥ 198,72 33 95 0,0861 TGF-β1 0,616 ≥ 8,21 43 100 0,0542		HVEM	0,916	≥ 1524,59	90	90	0,0000
TGF-β1 0,616 ≥ 8,21 43 100 0,0542	Other	Arginase	0,420	≥ 36,62	23	55	0,8537
		RANTES	0,600	≥ 198,72	33	95	0,0861
FAP 0,593 ≥ 141,20 33 100 0,1047		TGF-β1	0,616	≥ 8,21	43	100	0,0542
		FAP	0,593	≥ 141,20	33	100	0,1047

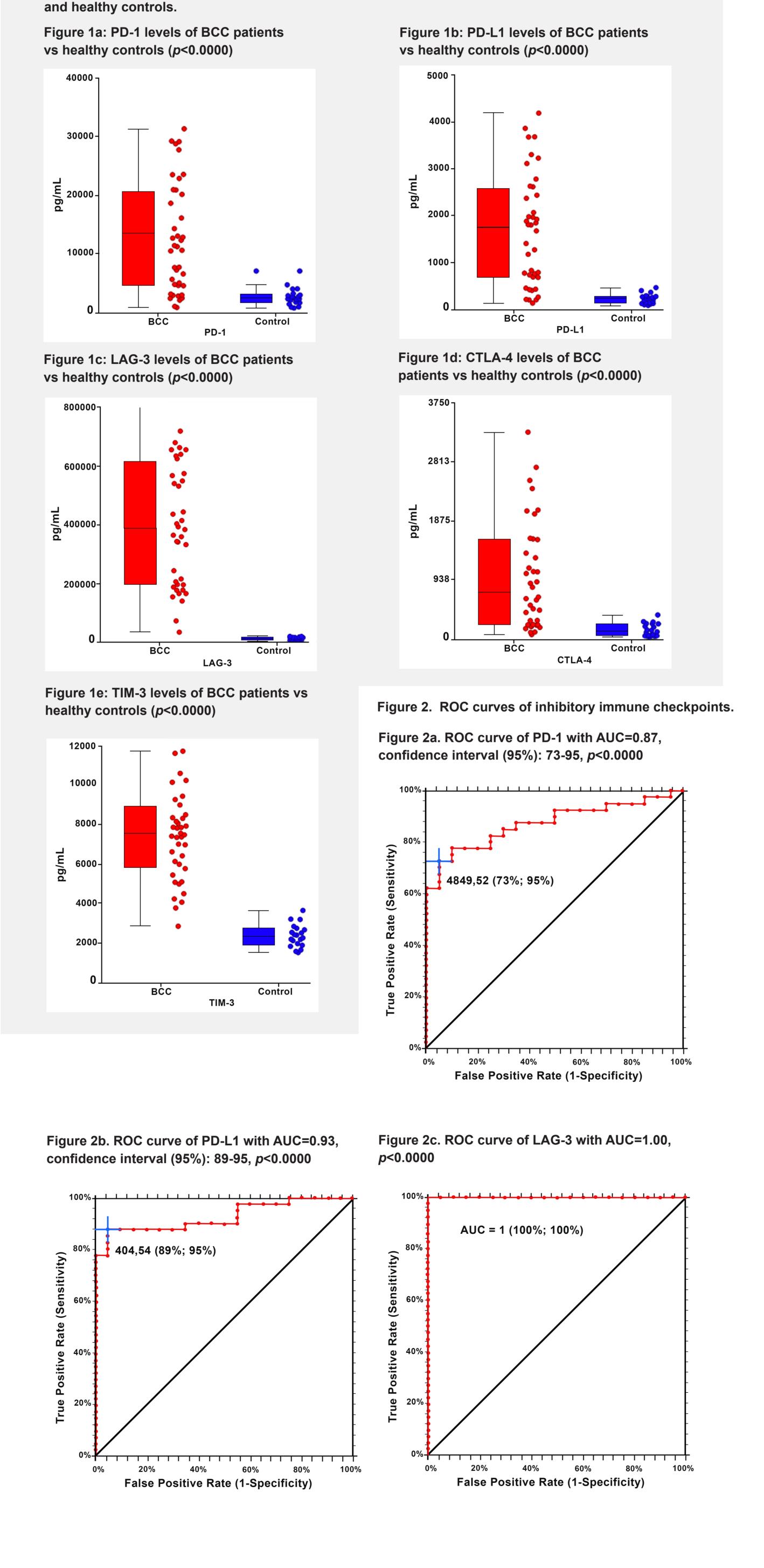
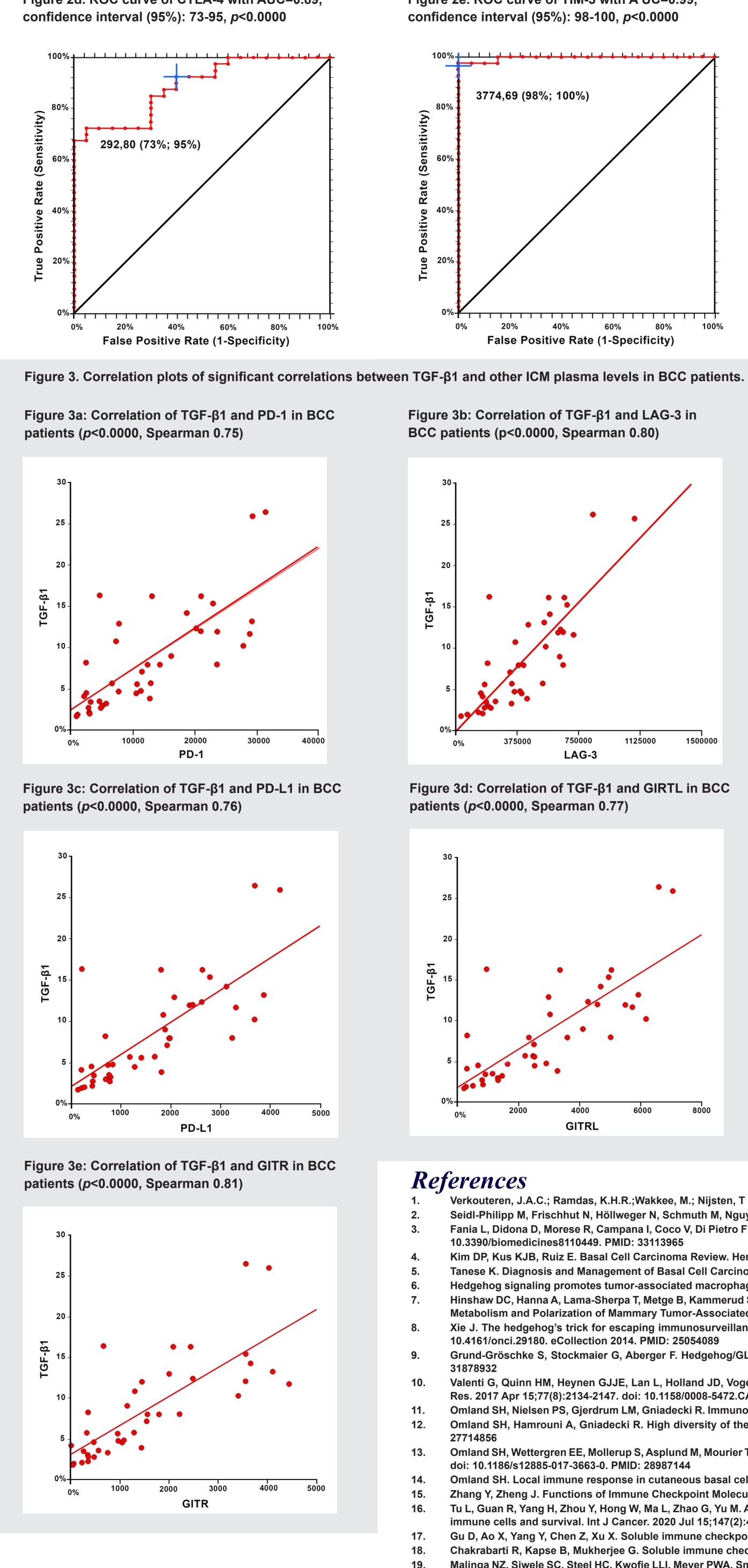
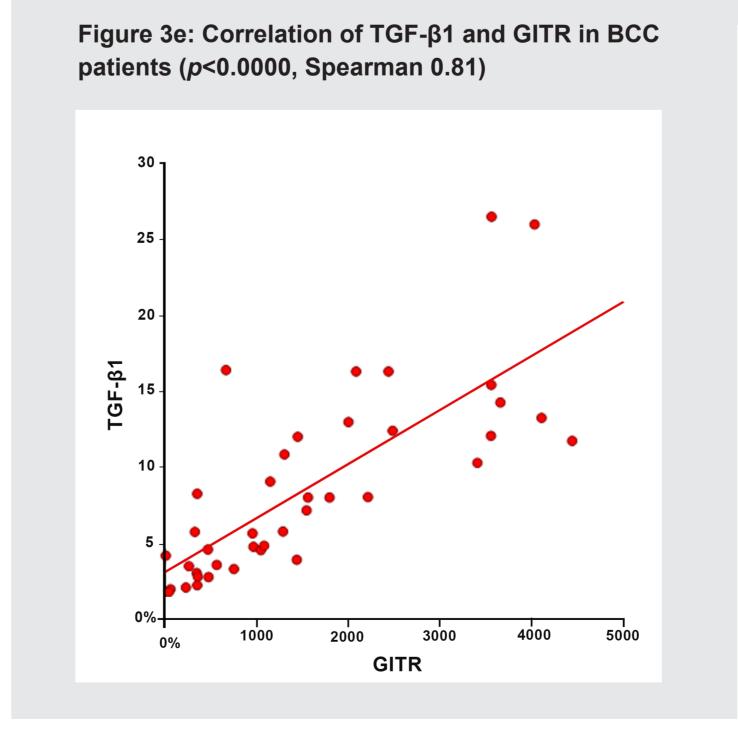
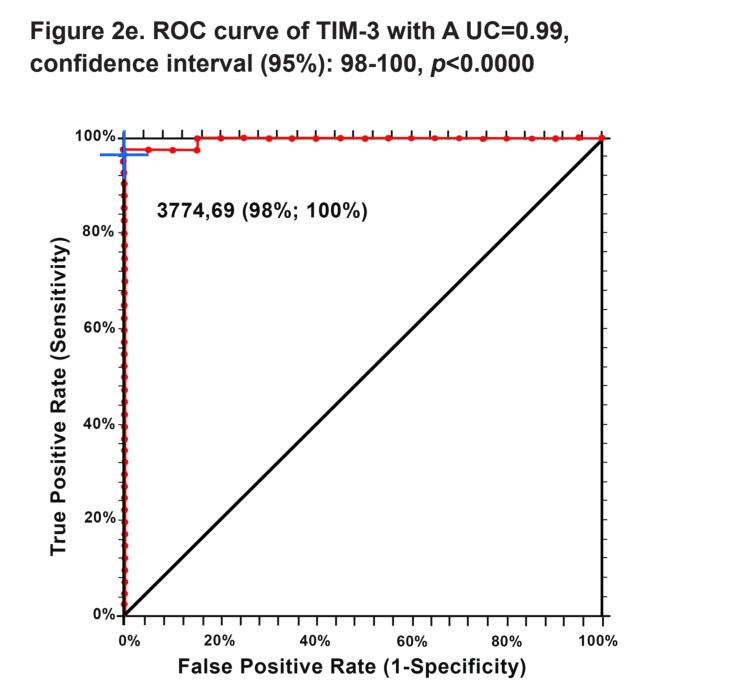
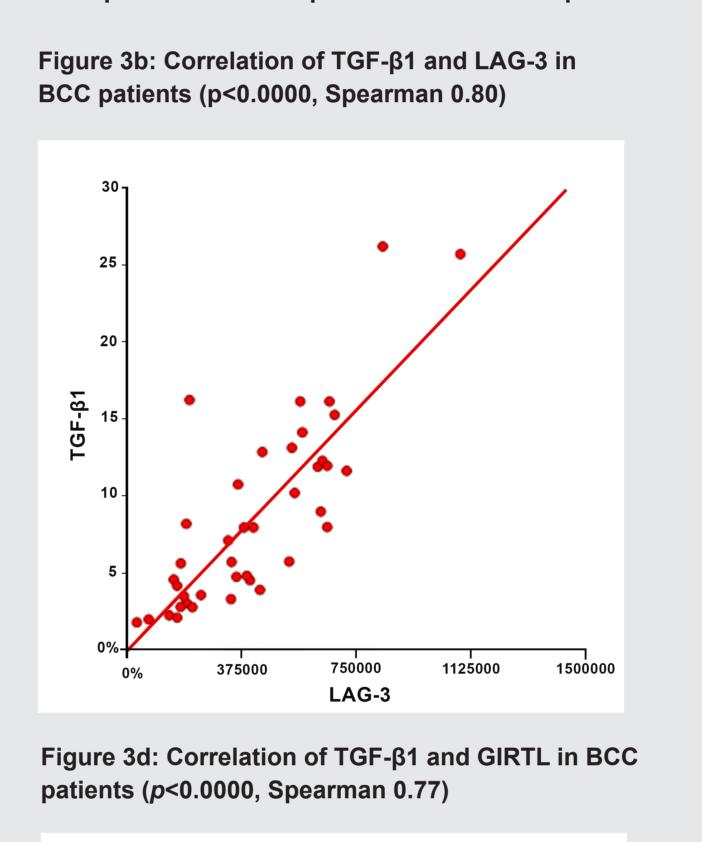


Figure 1. Comparison of plasma levels of inhibitory immune checkpoints between BCC patients









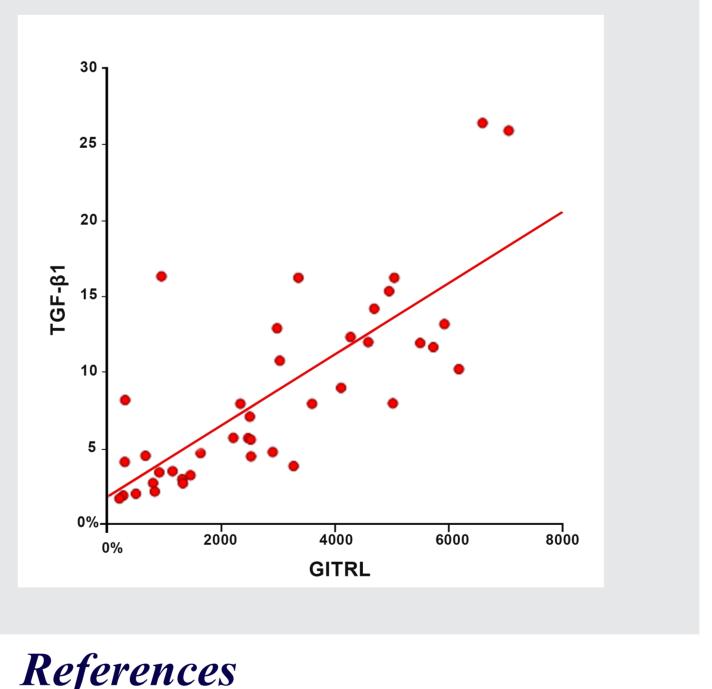
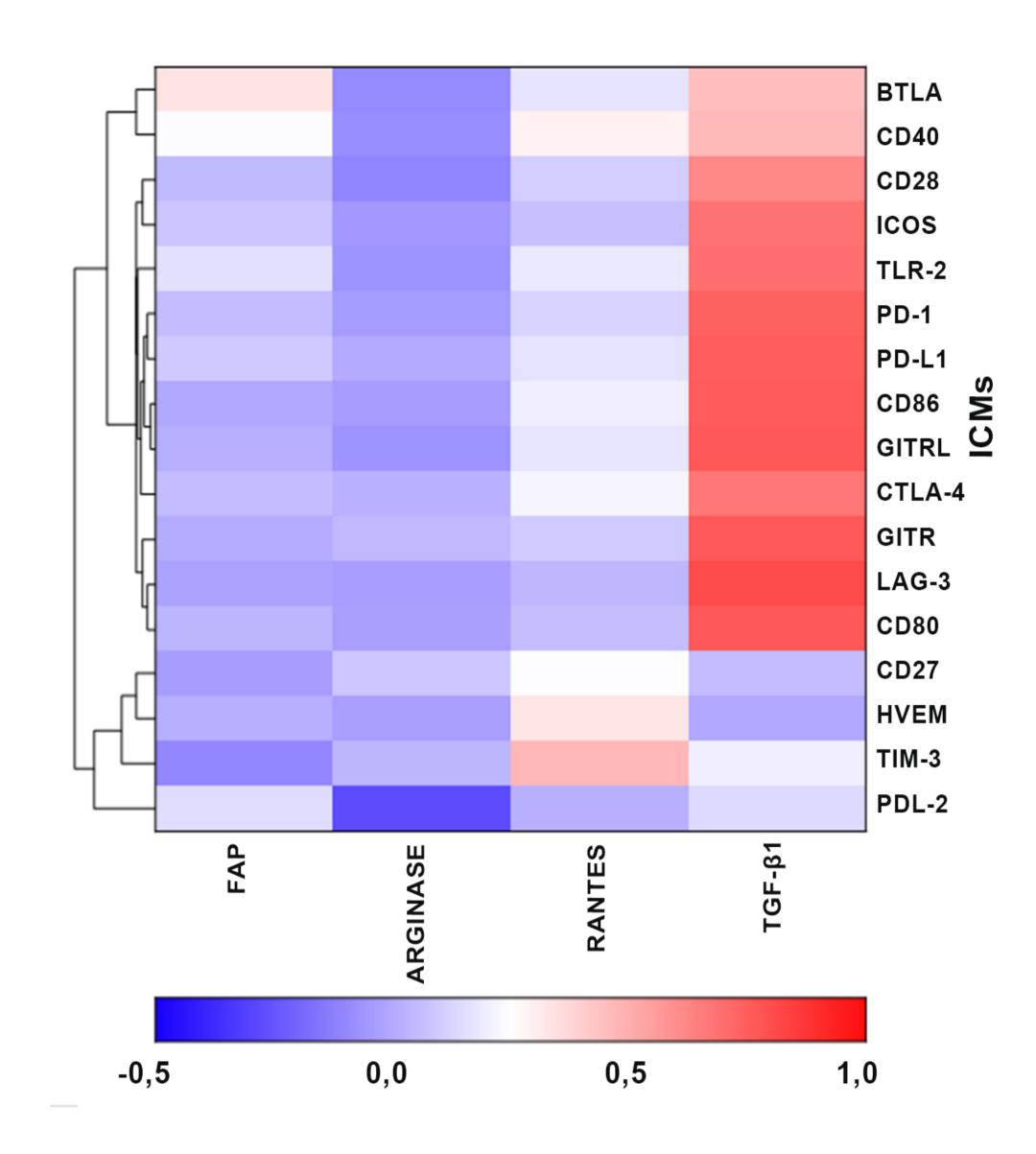


Figure 3. Clustered Heat Map.



Conclusions

- ▶ These seemingly novel findings not only identify the existence of significant systemic immunosuppression in BCC, but also underscore the therapeutic promise of immune checkpoint targeted therapy.
- ▶ The study demonstrates the potential of these proteins to serve as prognostic/predictive biomarkers in BCC.
- ▶ The therapeutic potential of triple targeting of PD-1 and TIM-3 or LAG-3 in this condition, as well as treatment with checkpoint inhibitors early in the course of the disease, is warranted.
- > We found plasma levels of TGF-β1, as a biomarker for Tregs, showing significant positive correlations with GITR, GITRL, LAG-3 PD-1, PD-L1, CD80, and CD86.
- ▶ There were no correlations found between any of the ICMs and FAP, arginase or RANTES respectively.

- Verkouteren, J.A.C.; Ramdas, K.H.R.; Wakkee, M.; Nijsten, T Epidemiology of basal cell carcinoma: scholarly review. Br J Dermatol. 2017 Aug; 177(2): 359-372. doi: 10.1111/bjd.15321. Epub 2017 Feb 20. PMID: 28220485 Seidl-Philipp M, Frischhut N, Höllweger N, Schmuth M, Nguyen VA. Known and new facts on basal cell carcinoma. J Dtsch Dermatol Ges. 2021 Jul;19(7):1021-1041. doi: 10.1111/ddg.14580. PMID: 34288482
- Fania L, Didona D, Morese R, Campana I, Coco V, Di Pietro FR, Ricci F, Pallotta S, Candi E, Abeni D, Dellambra E. Basal Cell Carcinoma: From Pathophysiology to Novel Therapeutic Approaches. Biomedicines. 2020 Oct 23;8(11):449. doi:
- Kim DP. Kus KJB. Ruiz E. Basal Cell Carcinoma Review. Hematol Oncol Clin North Am. 2019 Feb:33(1):13-24. doi: 10.1016/j.hoc.2018.09.004. PMID: 30497670
- Tanese K. Diagnosis and Management of Basal Cell Carcinoma. Curr Treat Options Oncol. 2019 Feb 11;20(2):13. doi: 10.1007/s11864-019-0610-0. PMID: 30741348 Review. Hedgehog signaling promotes tumor-associated macrophage polarization to suppress intratumoral CD8+ T cell recruitment. J Clin Invest. 2019 Dec 2;129(12):5151-5162. doi: 10.1172/JCl128644. PMID: 31638600 Hinshaw DC, Hanna A, Lama-Sherpa T, Metge B, Kammerud SC, Benavides GA, Kumar A, Alsheikh HA, Mota M, Chen D, Ballinger SW, Rathmell JC, Ponnazhagan S, Darley-Usmar V, Samant RS, Shevde LA. Hedgehog Signaling Regulates Metabolism and Polarization of Mammary Tumor-Associated Macrophages. Cancer Res. 2021 Nov 1;81(21):5425-5437. doi: 10.1158/0008-5472.CAN-20-1723. Epub 2021 Jul 21. PMID: 34289986.
- Xie J. The hedgehog's trick for escaping immunosurveillance: The molecular mechanisms driving myeloid-derived suppressor cell recruitment in hedgehog signaling-dependent tumors. Oncoimmunology. 2014 Jun 5;3:e29180. doi: Grund-Gröschke S, Stockmaier G, Aberger F. Hedgehog/GLI signaling in tumor immunity - new therapeutic opportunities and clinical implications. Cell Commun Signal. 2019 Dec 26;17(1):172. doi: 10.1186/s12964-019-0459-7. PMID:
- Valenti G, Quinn HM, Heynen GJJE, Lan L, Holland JD, Vogel R, Wulf-Goldenberg A, Birchmeier W. Cancer Stem Cells Regulate Cancer-Associated Fibroblasts via Activation of Hedgehog Signaling in Mammary Gland Tumors. Cancer Res. 2017 Apr 15:77(8):2134-2147. doi: 10.1158/0008-5472.CAN-15-3490. Epub 2017 Feb 15. PMID: 28202523
- Omland SH, Nielsen PS, Gjerdrum LM, Gniadecki R. Immunosuppressive Environment in Basal Cell Carcinoma: The Role of Regulatory T Cells. Acta Derm Venereol. 2016 Nov 2;96(7):917-921. doi: 10.2340/00015555-2440. PMID: 27117439 Omland SH, Hamrouni A, Gniadecki R. High diversity of the T-cell receptor repertoire of tumor-infiltrating lymphocytes in basal cell carcinoma. Exp Dermatol. 2017 May;26(5):454-456. doi: 10.1111/exd.13240. Epub 2017 Jan 19. PMID:
- Omland SH, Wettergren EE, Mollerup S, Asplund M, Mourier T, Hansen AJ, Gniadecki R. Cancer associated fibroblasts (CAFs) are activated in cutaneous basal cell carcinoma and in the peritumoural skin. BMC Cancer. 2017 Oct 7;17(1):675.
- Omland SH. Local immune response in cutaneous basal cell carcinoma. Dan Med J. 2017 Oct:64(10):B5412. PMID: 28975891 Zhang Y, Zheng J. Functions of Immune Checkpoint Molecules Beyond Immune Evasion. Adv Exp Med Biol. 2020;1248:201-226. doi: 10.1007/978-981-15-3266-5_9. PMID: 32185712
- 16. Tu L, Guan R, Yang H, Zhou Y, Hong W, Ma L, Zhao G, Yu M. Assessment of the expression of the immune checkpoint molecules PD-1, CTLA4, TIM-3 and LAG-3 across different cancers in relation to treatment response, tumor-infiltrating immune cells and survival. Int J Cancer. 2020 Jul 15;147(2):423-439. doi: 10.1002/ijc.32785. Epub 2019 Dec 2. PMID: 31721169
- Gu D, Ao X, Yang Y, Chen Z, Xu X. Soluble immune checkpoints in cancer: production, function and biological significance. J Immunother Cancer. 2018 Nov 27;6(1):132. doi: 10.1186/s40425-018-0449-0. PMID: 30482248 Chakrabarti R, Kapse B, Mukherjee G. Soluble immune checkpoint molecules: Serum markers for cancer diagnosis and prognosis. Cancer Rep (Hoboken). 2019 Aug;2(4):e1160. doi: 10.1002/cnr2.1160. Epub 2019 Feb 7.PMID: 32721130 Malinga NZ, Siwele SC, Steel HC, Kwofie LLI, Meyer PWA, Smit T, Anderson R, Rapoport BL, Kgokolo MCM. Systemic levels of the soluble co-inhibitory immune checkpoints, CTLA-4, LAG-3, PD-1/PD-L1 and TIM-3 are markedly increased in basal cell carcinoma, Transl Oncol. 2022 May:19:101384, doi: 10.1016/j.tranon.2022.101384, Epub 2022 Mar 4, PMID: 35255355